Abstract

Background: Lead is known for its adverse effects on various organs and systems. In this study, the ability of lead to adversely affect blood parameters was investigated, and *Lycopersicon esculentum*, or commonly known as tomato (a source of antioxidants), was administered orally in the form of tomato paste (TP) to reduce the adverse effects of lead.

Methods: The study involved 56 Wistar rats divided equally into 4 groups of 14 rats each: Control, LAG, tPG, and LA+tPG. Control and tPG rats were given distilled water ad libitum, while LAG and LA+tPG rats were given 1% lead (II) acetate (LA) per day. tPG and LA+tPG rats were additionally treated with 1.5 ml of TP per day. All treatments lasted for 10 weeks, after which the rats were weighed and sacrificed, and hematological and biochemical parameters were measured. The independent samples t test was used to analyse the results.

Results: Lead caused significant reductions in the following parameters: weight; packed cell volume; red blood cell and white blood cell counts; the percentages of lymphocytes and monocytes; total serum protein, albumin, and globulin levels; and plasma superoxide dismutase and catalase activities. In contrast, lead caused a significant increase in the percentage of neutrophils and the plasma malondialdehyde concentration. TP, however, significantly prevented the adverse effects of LA.

Conclusion: The oral administration of TP prevents the adverse effects of lead on blood constituents.

Keywords: anaemia, antioxidants, biochemical marker, lead, *Lycopersicon esculentum*, medical sciences

Introduction

Lead, a heavy metal, is harmful even in small amounts. Nevertheless, humans are exposed to lead through their environments and diets (1). Lead can be inhaled in dust from lead-based paints (2,3) or from fumes of leaded gasoline. It is found in trace amounts in various foods, notably in fish, which are heavily subjected to industrial pollution. Some old houses may have lead water pipes, which can easily contaminate drinking water (3). The manifestations of lead poisoning in humans are non-specific and may be hard to detect during the initial stages. Symptoms include weight loss, anaemia (4), memory loss (5), nephropathy, and infertility (6). Previous research has established that oxidation accompanies lead toxicity (7). This finding suggests that an ample source of antioxidants could reasonably prevent (or at least suppress) the manifestations of lead toxicity.

*Lycopersicon esculentum*, commonly known as tomato, is a good source of antioxidants (8). It contains nutrients that prevent illness, e.g., by detoxification (8,9), promoting growth (10) and proper immune system functioning (11), as well as increasing the haematocrit, RBC, and WBC content (4). Cooking helps to liberate the beneficial components of the tomato (lycopene, glutathione, vitamin C, and vitamin A) and also facilitates their absorption in the gastrointestinal tract (12,13). There is a possibility that oral administration of cooked tomatoes could reasonably prevent (or at least suppress) the manifestations of lead toxicity. Therefore, this study focused on the potential of oral administration of cooked tomatoes in preventing or suppressing lead toxicity.

Materials and Methods

Animals

A total of 56 adult male Sprague–Dawley rats (average body weight 183.45 ± 0.903 g) were used for this study. They were inbred at the Animal House of the Faculty of Basic Medical Sciences, Ladoke Akintola University of Technology, Ogbomoso, Nigeria. The rats were transferred to the research section and acclimatised over a period of 2 weeks. The research section was maintained...
at a temperature of 24–26 °C, relative humidity of 70%–75%, and a light/dark cycle of 12/12 hours with adequate ventilation throughout the period of research. The rats were fed with commercially available rat pellets. The maintenance, care, and treatments of the rats complied with National Institutes of Health guideline for the humane use of laboratory rats and the guideline of Ladoke Akintola University of Technology.

Preparation of tomato paste
Since no published method is available for concentrating the tomato paste, the following in-house method was adopted. Furthermore, cooking has been shown to liberate the beneficial/active components of tomatoes (lycopene, glutathione, vitamin C, and vitamin A) and facilitate their absorption in the gastrointestinal tract (12,13).

Tomato paste (TP) was prepared by grinding tomatoes and heating them in a water bath at 80°C for 45 to 60 minutes. The obtained TP was removed from the water bath when its relative density was 1.124. It was then allowed to cool to 25 °C, and then stored at 4 °C. The quantity of TP to be administered was thawed to 25 °C 1 hour prior to administration.

Treatments
The rats were randomly divided into 4 groups: 1) control given distilled water, 2) LA group given 1% lead (II) acetate (LA), 3) TPG group given distilled water and 1.5 ml of TP per day, and 4) LA+TPG group given 1% lead (II) acetate (LA) and 1.5 ml of TP per day. All groups consisted of 14 rats each. All treatments lasted 10 weeks.

Sample collection and processing
Each animal was weighed and sacrificed by cervical dislocation 24 hours after the last treatment. Blood samples were collected via cardiac puncture. The blood obtained from each rat was injected into 2 tubes; a plain (no-additive) tube and an EDTA tube. The blood samples in the EDTA tubes were mixed thoroughly (although gently) with the EDTA. All the blood samples were then stored below room temperature (10–14 °C) prior to the measurement of haematological and biochemical parameters.

Measurement of haematological and biochemical parameters and data analysis
Packed cell volume (PCV) was determined using plain capillary tubes filled with anticoagulated blood and centrifugation at 3000 rpm for 20 minutes. Red blood cell (RBC) and white blood cell (WBC) counts were determined using an improved Neubauer counting chamber following the procedure documented by Cheesbrough and McArther (14). Field’s stain A and stain B were used for the differential WBC counts. Plasma and serum were obtained by centrifugation of whole blood at 3000 rpm for 20 minutes. To serve as a guide, total serum protein (TSP), total serum albumin (TSA), and total serum globulin (TSG) levels were first estimated using the Levibond comparator (i.e., the Biuret method) following the method documented by Cheesbrough and McArther (14), and accurate (reported) values for TSP, TSA, and TSG were then determined using photo-electric colourimeter. The total plasma protein (TPP) was on the other hand determined using the following formula derived by Kagan (15):

\[ P = 340(G - 1.0099) \]

where P is the total grams of protein per 100 cm³ of plasma and G is the specific gravity of plasma at 25 °C.

Plasma superoxide dismutase (SOD) activity was determined using the method described by Fridovich (16). Plasma catalase (CAT) activity was determined using the method described by Sinha (17). Plasma malondialdehyde (MDA) concentrations were determined using the procedure described by Varshney and Kale (18). SOD, CAT, and MDA were chosen because many previous studies have reported their reliability in the measurement of antioxidant activities and level of lipid peroxidation (19–22).

The data obtained are presented as mean ± SEM. The control and the treatment groups were compared using the independent samples t test using SPSS version 14 (SPSS Inc, Chicago, IL). The level of significance was set at a P < 0.05.

Results
Weight gain
Comparison of the initial and final weights of rats showed that there was significant weight gain (P < 0.05) in all the groups at the end of 10 weeks of treatment (Table 1). There was no significant difference in weight gain in the TP, LA+TP, and control groups. However, the increase of weight in the LA group was significantly lower than that of the control group (P < 0.05).

Haematological parameters
Haematological analyses indicated that LA rats (unlike TP and LA+TP rats) have significantly lower PCV, RBC count, and WBC count (P < 0.01 each) (Table 2), as well as lower
Original Article | *Lycopersicon esculentum* preserves blood constituents

Table 1: Weight of rats before and after treatments

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>LA&lt;sub&gt;G&lt;/sub&gt;</th>
<th>TP&lt;sub&gt;G&lt;/sub&gt;</th>
<th>LA+TP&lt;sub&gt;G&lt;/sub&gt;</th>
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<tbody>
<tr>
<td>Initial weight (g)</td>
<td>183.2 ± 1.02</td>
<td>182.1 ± 0.34</td>
<td>184.0 ± 0.54</td>
<td>184.5 ± 0.44</td>
</tr>
<tr>
<td>Weight after treatment (g)</td>
<td>219.3 ± 0.99</td>
<td>198.9 ± 0.54</td>
<td>221.2 ± 0.21</td>
<td>218.5 ± 0.32</td>
</tr>
<tr>
<td>Weight increase (g)</td>
<td>36.1 ± 0.33</td>
<td>16.8 ± 0.99</td>
<td>37.2 ± 0.76</td>
<td>34.0 ± 0.94</td>
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</table>

Control and TP<sub>G</sub> rats were given distilled water ad libitum, while LA<sub>G</sub> and LA+TP<sub>G</sub> rats were given 1% lead (II) acetate (LA) ad libitum daily. TP<sub>G</sub> and LA+TP<sub>G</sub> rats were also given 1.5 ml of TP per day. All treatments lasted 10 weeks. Data are expressed in mean ± SEM. *P* < 0.05 indicating significant difference when compared with respective initial weight by paired samples *t* test. *P* < 0.05 indicating significant difference when compared with the control group by independent samples *t* test.

Table 2: Packed cell volume (PCV), red blood cell (RBC) count, and white blood cell (WBC) count in control and treated groups

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>LA&lt;sub&gt;G&lt;/sub&gt;</th>
<th>TP&lt;sub&gt;G&lt;/sub&gt;</th>
<th>LA+TP&lt;sub&gt;G&lt;/sub&gt;</th>
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<tbody>
<tr>
<td>PCV (%)</td>
<td>33.6 ± 0.31</td>
<td>30.2 ± 0.24</td>
<td>34.1 ± 0.46</td>
<td>32.6 ± 0.25</td>
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<tr>
<td>RBC (thousand cells/mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>6334.0 ± 3.75</td>
<td>5358.0 ± 26.31 **</td>
<td>7431.3 ± 4.47</td>
<td>5867.3 ± 46.84</td>
</tr>
<tr>
<td>WBC (cells/mm&lt;sup&gt;3&lt;/sup&gt;)</td>
<td>2522.0 ± 3.02</td>
<td>2140.0 ± 2.14 **</td>
<td>2437.0 ± 2.46</td>
<td>2340.0 ± 2.46</td>
</tr>
</tbody>
</table>

Control and TP<sub>G</sub> rats were given distilled water ad libitum, while LA<sub>G</sub> and LA+TP<sub>G</sub> rats were given 1% lead (II) acetate (LA) ad libitum daily. TP<sub>G</sub> and LA+TP<sub>G</sub> rats were also given 1.5 ml of TP per day. All treatments lasted 10 weeks. Data are expressed in mean ± SEM. *P* < 0.05 and **P* < 0.01 indicated significant difference when compared with the control group by independent samples *t* test.

Discussion

The results showed that chronic exposure to lead significantly reduced weight gain, which support the findings of Wadi and Ahmad (23) and can be linked to less-efficient metabolic processes due to lead toxicity (24). Administration of 1.5 ml TP/day, however, eliminates the adverse effect of lead on weight gain. This result would be chiefly due to presence of protective antioxidants in TP, such as lycopene, vitamin C, and vitamin A (7), even though it has low protein content and low caloric value (18 kcal/100 g). Similar reasons could be attributed to the significant decrease in PCV observed in rats exposed to lead (LAG) and the non-significant difference in PCV observed in rats treated with TP and lead (LA+TP<sub>G</sub>) when compared with the control group. The finding that lead decreased PCV agrees with the findings of Anetor et al. (25) that some indices of erythropoietic activity, such as haemoglobin (Hb) concentration, packed cell volume (PCV), and mean corpuscular haemoglobin concentration (MCHC), were significantly decreased in workers exposed to lead compared to the control group. However, Franson et al. (26) reported that PCV is not altered in birds after supplementation of 50 ppm of lead in their feed, and Arvind and Chopra

percentages of lymphocytes and monocytes (*P* < 0.05 each), but significantly higher percentages of neutrophils (*P* < 0.05) when compared with the control group (Table 3). The percentages of eosinophils and basophils for each of the groups were, however, not significantly different from those of the control group.

Protein content

TSP, TSA, and TSG levels for LA<sub>G</sub> rats were significantly lower (*P* < 0.05) than those of control group, while the levels in TP<sub>G</sub> and LA+TP<sub>G</sub> rats were not significantly different from the control group (Table 4).

Antioxidant activities and lipid peroxidation

LA<sub>G</sub> rats had significant decreases in plasma SOD and CAT activities (*P* < 0.01); whereas TP<sub>G</sub> animals had significantly higher SOD (*P* < 0.01) and CAT (*P* < 0.05) activities when compared with the control group (Table 5). LA<sub>G</sub> rats had significant increases in plasma MDA concentration (*P* < 0.05), while TP<sub>G</sub> had significant decreases in MDA concentration (*P* < 0.05) compared with the control group. On the other hand, LA+TP<sub>G</sub> rats had no significant differences in SOD activity, CAT activity, and MDA concentration compared with the control group.
Table 3: White blood cell (WBC) differential count in control and treated groups

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>LAG</th>
<th>TP</th>
<th>LA+TP</th>
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<tbody>
<tr>
<td>Neutrophils (%)</td>
<td>58.4 ± 0.29</td>
<td>62.4 ± 0.19*</td>
<td>56.4 ± 0.21</td>
<td>58.4 ± 0.31</td>
</tr>
<tr>
<td>Lymphocytes (%)</td>
<td>31.2 ± 0.24</td>
<td>29.6 ± 0.15*</td>
<td>32.0 ± 0.43</td>
<td>31.0 ± 0.27</td>
</tr>
<tr>
<td>Monocytes (%)</td>
<td>6.4 ± 0.25</td>
<td>4.6 ± 0.15*</td>
<td>7.1 ± 0.12</td>
<td>6.0 ± 0.22</td>
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<tr>
<td>Eosinophils (%)</td>
<td>3.2 ± 0.13</td>
<td>2.8 ± 0.18</td>
<td>3.3 ± 0.34</td>
<td>3.6 ± 0.23</td>
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<tr>
<td>Basophils (%)</td>
<td>0.8 ± 0.13</td>
<td>0.6 ± 0.15</td>
<td>1.2 ± 0.24</td>
<td>1.0 ± 0.17</td>
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</table>

Control and TP<sub>G</sub> rats were given distilled water ad libitum, while LAG and LA+TP<sub>G</sub> rats were given 1% lead (II) acetate (LA) ad libitum daily. TP<sub>G</sub> and LA+TP<sub>G</sub> rats were also given 1.5 ml of TP per day. All treatments lasted 10 weeks. Data are expressed in mean ± SEM. *P < 0.05 indicated significant difference when compared with the control group by independent samples t test.

Table 4: Total serum protein (TSP), total serum albumin (TSA), and total serum globulin (TSG) levels in the control and treated groups

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>LAG</th>
<th>TP</th>
<th>LA+TP</th>
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<tbody>
<tr>
<td>TSP (g/L)</td>
<td>74.8 ± 0.36</td>
<td>63.2 ± 0.51*</td>
<td>76.7 ± 0.36</td>
<td>70.8 ± 0.26</td>
</tr>
<tr>
<td>TSA (g/L)</td>
<td>45.6 ± 0.49</td>
<td>39.4 ± 0.38*</td>
<td>47.1 ± 0.23</td>
<td>42.4 ± 0.27</td>
</tr>
<tr>
<td>TSG (g/L)</td>
<td>29.2 ± 0.41</td>
<td>23.8 ± 0.423*</td>
<td>29.6 ± 0.09</td>
<td>28.4 ± 0.25</td>
</tr>
</tbody>
</table>

Control and TP<sub>G</sub> rats were given distilled water ad libitum, while LAG and LA+TP<sub>G</sub> rats were given 1% lead (II) acetate (LA) ad libitum daily. TP<sub>G</sub> and LA+TP<sub>G</sub> rats were also given 1.5 ml of TP per day. All treatments lasted 10 weeks. Data are expressed in mean ± SEM. *P < 0.05 indicated significant difference when compared with the control group by independent samples t test.

Table 5: Plasma superoxide dismutase (SOD) activity, catalase (CAT) activity, and malondialdehyde (MDA) concentration in control and treated groups

<table>
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<tr>
<th></th>
<th>Control</th>
<th>LAG</th>
<th>TP</th>
<th>LA+TP</th>
</tr>
</thead>
<tbody>
<tr>
<td>SOD activity</td>
<td>1.77 ± 0.05</td>
<td>1.12 ± 0.06</td>
<td>2.00 ± 0.19</td>
<td>1.70 ± 0.09</td>
</tr>
<tr>
<td>CAT activity</td>
<td>0.41 ± 0.08</td>
<td>0.22 ± 0.03</td>
<td>0.44 ± 0.04</td>
<td>0.40 ± 0.09</td>
</tr>
<tr>
<td>MDA concentration</td>
<td>1400.3 ± 23.01</td>
<td>1813.6 ± 11.18*</td>
<td>1152.5 ± 31.43*</td>
<td>1419.5 ± 34.07*</td>
</tr>
</tbody>
</table>

Control and TP<sub>G</sub> rats were given distilled water ad libitum, while LAG and LA+TP<sub>G</sub> rats were given 1% lead (II) acetate (LA) ad libitum daily. TP<sub>G</sub> and LA+TP<sub>G</sub> rats were also given 1.5 ml of TP per day. All treatments lasted 10 weeks. Data are expressed in mean ± SEM. *P < 0.05 and **P < 0.01 indicated significant difference when compared with the control group by independent samples t test.

(27) also reported no changes in PCV in calves after supplementation of 100 ppm of lead in their diets. This may considerably be due to the low concentration of lead (50 ppm or 0.005%, and 100 ppm or 0.010%, respectively) administered, and/or the different species used in the experiments.

There was no significant decrease in RBC counts in animals co-treated with TP and lead. In contrast, the RBC counts in LAG<sub>G</sub> rats were significantly lower than those of control group. The reduced in RBC counts may be caused by lead interference in the energy metabolism of erythrocytes (28), which affects the ATP concentration in erythrocytes, thus, shortening their life span and lowering the blood counts. Morse et al. (29) reported that acute lead toxicity in mice produced transient erythroid hypoplasia and impaired utilisation of RBC ⁵⁹Fe for haem synthesis. The administered TP would therefore
be responsible for preventing these effects of lead on RBC probably by preventing the adverse effects of lead on the energy metabolism of erythrocytes and the utilisation of RBC $^{59}$Fe for haem synthesis and/or by elevating RBC glutathione level (a function of vitamin C component in tomato), which protects RBC against damaged caused by hydrogen peroxide (a toxic by-product of many metabolic reactions) by reducing the peroxide to water (30).

In a similar manner, WBC counts in rats co-treated with TP and lead were not significantly different from those of control group. Conversely, rats treated with lead had significant decreases in WBC counts. This result suggests that TP combats the adverse effects of lead on WBC, resulting in no significant differences in WBC counts observed in control group and in rats co-treated with TP and lead. This finding was most likely due to the detoxification effect of some antioxidant components of TP. Similar detoxification/protective effects have previously been documented for varieties of antioxidants, such as carotene, retinol, and bioflavonoids (31, 32).

Lead caused significant decreases in lymphocyte and monocyte counts but increased neutrophil counts. These results support the finding by Sembulingam and Sembulingam (33) that lead (like some other chemicals and drugs, e.g., mercury, camphor, benzene derivatives, venoms, and some vaccines) causes neutrophilia. Di Lorenzo et al. (34) also reported that mean absolute neutrophil counts were significantly higher in workers exposed to lead compared to workers who were not exposed to lead. TP prevented lead-induced changes to WBC and differential counts. The percentages of eosinophils and basophils in rats exposed to lead alone and those of rats treated with TP after lead exposure were not significantly different from those of control group.

The significantly lower TSP, TSA, and TSG levels in rats exposed to lead could result from the fact that lead causes disturbances in metabolism, reduces the efficiency of gastrointestinal tract digestion and absorption processes, and negatively affects protein synthesis (4,25). TP, in contrast, was able to annul the adverse effects of lead on TSP, TSA, and TSG levels of rats co-treated with TP and lead. This effect was perhaps due to some protein (sources of amino acids) components of TP (which might be used as raw materials for, or facilitate, aminogenesis/proteogenesis) and could, better still, be traced to its vitamins and lycopene components and TP ability to modulate gastrointestinal tract digestive and absorptive functions (9).

There were significant decreases in plasma SOD and CAT activities in lead-treated rats, while in rats co-treated with TP and lead, no significant differences were observed in comparison with the control group. These findings are in agreement with those of Hsu and Guo (35). Although plasma MDA concentration was significantly increased in lead-treated rats, no significant difference was observed in rats co-treated with TP and lead in comparison with the control group. This confirms that TP, as a source of antioxidants (7, 8), reduced the oxidative stress caused by lead exposure in this animal model.

**Conclusion**

Exposure to lead significantly reduced weight gain; PCV, RBC count, and WBC count; percentages of lymphocytes and monocytes; TSP, TSA, and TSG levels; and plasma SOD and CAT activities. In contrast, it caused significant increases in the percentages of neutrophils and eosinophils, and plasma MDA concentration. TP, however, significantly reduced these adverse effects of lead. It would be good if subsequent studies could examine if human consumption of considerable (high) amount of TP could have protective effect against lead toxicity, and if prolonged consumption of TP has any potential adverse effects in humans.

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