Bacterial isolates and drug susceptibility patterns of urinary tract infection among pregnant women at Muhimbili National Hospital in Tanzania

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Abstract: Urinary tract infection (UTI) during pregnancy may cause complications such as pyelonephritis, hypertensive disease of pregnancy, anaemia, chronic renal failure, premature delivery and foetal mortality. This study aimed to identify the etiologic agents of UTI and to determine the patterns of antimicrobial drug susceptibility among pregnant women at Muhimbili National Hospital in Tanzania. Retrospective analysis of 200 mid-stream urine specimens processed for culture and antimicrobial drug susceptibility testing between January 2007 and December 2009 was carried out. Significant bacteriuria (> 10^5 colony forming units/mL of urine) was found in 42/200 (21%) specimens. Of the 42 isolates, the most commonly isolated bacteria were Escherichia coli 14 (33.3%), Klebsiella spp 9 (21.4%) coagulase negative Staphylococcus 7 (16.7%), Staphylococcus aureus 6 (14.3%), Proteus species 3 (7.1%) and Enterococcus species 3 (7.1%). Low rate to moderately high rate of antimicrobial drug resistance was observed against first line drugs namely, nitrofurantoin 18.7 % (n=16), co-trimoxazole 38.5 % (n=13) and ampicillin 57.7 % (n=26). Relatively low rate of resistance was seen against second line drugs: ciprofloxacin 13.6 % (n=22) and amikacin 5 % (n=20). High rate of resistance was observed in third generation cephalosporin cefotaxime 31.2 % (n=16). Of the Gram-positive organisms tested against vancomycin and mexitelincin, resistance was found in 25 % (n=13) and 25 % (n=4), respectively. In conclusion, E coli was found to be the common cause of UTI among the pregnant women. Low to moderately high level of resistance was found in first line drugs while high level of resistance was found in third generation cephalosporin. It is recommended to monitor the levels of resistance for nitrofurantoin, fluoroquinolone and cefotaxime and to screen for Extended Spectrum Beta Lactamase production among cefotaxime resistant E. coli and Klebsiella spp.

Key words: Urinary tract infection, pregnant women, antimicrobial drug resistance, Tanzania

Introduction

Urinary tract infection (UTI) is a common health problem among pregnant women (Mittal & Wing, 2005; Saidi et al., 2005). This usually begins in week 6 and peaks during weeks 22 to 24 of pregnancy due to a number of factors including ureteral dilatation, increased bladder volume and decreased bladder tone, along with decreased ureteral tone which contributes to increased urinary stasis and ureterovesical reflux (Chaliha et al., 2002). Up to 70% of pregnant women develop glycosuria, which encourages bacterial growth in the urine (Al-Issa, 2009).

UTI may manifest as asymptomatic bacteriuria (ASB) or symptomatic bacteriuria (SB). The prevalence of asymptomatic UTI has been previously reported to be 2% to 13% in pregnant women (Delzell et al., 2000; Christensen, 2000; Kutlay et al., 2003; McIsaac, 2005; Masinde et al., 2009) compared with that of symptomatic UTI which occurs in 1–18% during pregnancy (Dwyer & O’Reilly, 2002; Masinde et al., 2009).

UTI during pregnancy may cause complications such as pyelonephritis, hypertensive disease of pregnancy, anaemia, chronic renal failure, premature delivery and foetal mortality (Delzell et al., 2000; Foxman, 2002). The incidence of these complications can be decreased by treating promptly ASB and SB during pregnancy (Delzell et al., 2000). Due to the potential adverse sequelae of UTI in pregnancy, most clinics perform routine...
urinalysis of midstream urine specimen during one or more antenatal clinic (ANC) visits (Smaill, 2007). However, culture and antimicrobial drug susceptibility testing are needed for surveillance purposes to guide the clinicians on the proper management and prevent empirical treatment of pregnant women with ASB and SB.

A limited spectrum of organisms cause UTI and these include *Escherichia coli*, which accounts for the majority of uncomplicated UTI isolates (Gupta et al., 2001; Nicolle, 2001; Masinde et al., 2009). Others are *Staphylococcus saprophyticus*, *Klebsiella* spp, *Proteus* spp, *Enterococcus* spp, and *Enterobacter* spp (Gupta et al., 2001; Masinde et al., 2009). Data on the current distribution and antimicrobial drug susceptibility patterns among urinary bacterial isolates from pregnant women in Tanzania is limited. This study aimed to identify the etiologic agents of UTI and to determine the patterns of antimicrobial drug susceptibility among pregnant women at Muhimbili National Hospital (MNH), Dar es Salaam, Tanzania.

**Material and methods**

**Study area**

A retrospective review and analysis of 3-year laboratory records (2007-2009) on urine culture specimens from pregnant women attending ANC at Muhimbili National Hospital in Tanzania was carried out. Data were extracted from laboratory registers in the Microbiology Section at Central Pathology Laboratory (CPL) of the Hospital. Information on the type of bacterial isolates from the urine specimens and antibiotic susceptibility patterns were recorded.

**Bacterial isolation and antimicrobial susceptibility testing**

Urine samples received at the Central Pathology Laboratory (CPL) were plated on cysteine lactose electrolyte deficient (CLED) agar medium incubated at 37°C for 24 hours. Growth of one type of organism of >10^5 colony forming units was considered as significant bacteriuria. Identification of isolates was done by observing colonial morphology on CLED medium. Colonies were further identified using standard biochemical tests as described previously (McCartney et al., 1989).

Kirby-Bauer disk diffusion test was used to perform antimicrobial susceptibility testing for all the isolates as recommended by Clinical and Laboratory Standard Institute (CLSI 2006). Antibiotic disks (Oxoid) were applied to each plate. Inhibition zone sizes were measured and interpreted according to CLSI guidelines. Antimicrobial drug susceptibility testing for cotrimoxazole, ampicillin, ciprofloxacin, amikacin, nitrofurantoin, methicillin, vancomycin and cefotaxime was done on all bacteria isolated.

**Results**

During the period of 2007-2009, a total of 200 urine specimens was received from ANC and processed. Significant bacteriuria (cultures with > 10^5 colony forming units (cfu) of bacteria/mL of urine) was found in 42/200 (21%; range=18.7-23.2%) of the urine specimens. Mixed bacterial growths were observed in 14 specimens (Table 1). Of all the bacteria isolated, Gram-negative bacteria were more prevalent (61.9%) than Gram-positive bacteria which constituted 38.1%. The most commonly isolated bacteria were *Escherichia coli* 14 (33.3%), *Klebsiella* species 9 (21.4%), coagulase negative *Staphylococcus* 7 (16.7%), *S. aureus* 6 (14.3%), *Proteus* species 3 (7.1%) and *Enterococcus* species 3 (7.1%).

<table>
<thead>
<tr>
<th>Organism isolated</th>
<th>2007</th>
<th>2008</th>
<th>2009</th>
<th>Total (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Escherichia coli</em></td>
<td>4</td>
<td>7</td>
<td>3</td>
<td>14 (33.3)</td>
</tr>
<tr>
<td><em>Klebsiella</em> species</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>9 (21.4)</td>
</tr>
<tr>
<td><em>Proteus</em> species</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>3 (7.1)</td>
</tr>
<tr>
<td><em>S. aureus</em></td>
<td>4</td>
<td>0</td>
<td>2</td>
<td>6 (14.3)</td>
</tr>
<tr>
<td>Coagulase negative staphylococcus</td>
<td>3</td>
<td>2</td>
<td>2</td>
<td>7 (16.7)</td>
</tr>
<tr>
<td><em>Enterococcus</em> species</td>
<td>3</td>
<td>0</td>
<td>0</td>
<td>3 (7.1)</td>
</tr>
<tr>
<td>Total number of urine specimen received</td>
<td>91</td>
<td>56</td>
<td>53</td>
<td>200</td>
</tr>
<tr>
<td>Total positive urine culture</td>
<td>17</td>
<td>13</td>
<td>12</td>
<td>42</td>
</tr>
</tbody>
</table>

Table 1: Distribution of bacteria isolated from pregnant women presenting with symptoms of UTI at MNH, Dar es Salaam, Tanzania, from 2007 to 2009.
Overall bacterial sensitivity to first line antibiotics for UTI was: nitrofurantoin 81.3 % (n=16), ampicillin 42.3% (n=26) and co-trimoxazole 61.5 % (n=13). Sensitivity to second line drugs was: ciprofloxacin 86.4 % (n=22) and amikacin 95 % (n=20). Gram negative bacteria showed a 31.2% (n=16) resistance to third generation cephalosporin cefotaxime (another second line antibiotic for UTI). Of the Gram-positive organisms tested against vancomycin and methicillin, antibiotic resistance was found in 25 % (n=13) and 25 % (n=4), respectively of the isolates (Table 2).

### Table 2: Patterns of antimicrobial sensitivity results for the bacterial isolated from pregnant women presenting with UTI at MNH, Dar es Salaam, Tanzania, 2007-2009.

<table>
<thead>
<tr>
<th>Antimicrobial agents (number of isolates tested, n)</th>
<th>Sensitive isolates (S)</th>
<th>Resistant isolates (R)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sensitivity patterns for first line antimicrobial agents</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ampicillin (n=26) *</td>
<td>11 (42.3%)</td>
<td>15 (57.7%)</td>
</tr>
<tr>
<td>Cotrimoxazole(n=13) *</td>
<td>8 (61.5%)</td>
<td>5 (38.5%)</td>
</tr>
<tr>
<td>Nitrofurantoin (n=16) **</td>
<td>13 (81.3%)</td>
<td>3 (18.7%)</td>
</tr>
<tr>
<td><strong>Sensitivity patterns for second line antimicrobial agents</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amikacin (n=20) *</td>
<td>19 (95%)</td>
<td>1 (5%)</td>
</tr>
<tr>
<td>Ciprofloxacin (n=22) *</td>
<td>19 (86.4%)</td>
<td>3 (13.6%)</td>
</tr>
<tr>
<td>Cefotaxime (n=16) **</td>
<td>11 (68.8%)</td>
<td>5 (31.2%)</td>
</tr>
<tr>
<td><strong>Sensitivity pattern for gram positive cocci</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Methicillin (n=4)****</td>
<td>3 (75%)</td>
<td>1 (25%)</td>
</tr>
<tr>
<td>Vancomycin (n=13) ***</td>
<td>12 (95%)</td>
<td>1 (25%)</td>
</tr>
</tbody>
</table>

* tested against gram positive and gram negative bacteria; ** tested against gram negative only; *** tested against gram positive only; ****tested against Staphylococcus spp only

### Discussion

Proper investigation and prompt treatment are needed to prevent serious life threatening condition and morbidity due to UTI that can occur in pregnant women. Our study findings showed that only 200 (0.27%) urine specimens out of 73,894 antenatal clinic attendees were tested for culture and antibiotic susceptibility patterns during the last three years. The small number of specimens may be explained by the fact that only those suspected to have UTI were investigated. We recommend screening of all pregnant women for significant bacteriuria because ASB can be present in 2% to 13% of pregnant women (Delzell et al., 2000; Kutlay et al. 2003; McIsaac et al., 2005; Masinde et al., 2009). Moreover, previous clinical trials have shown that antibiotic treatment significantly reduces the risk of complications associated with UTI in pregnancy. Furthermore, screening for and treatment of asymptomatic bacteriuria during pregnancy has become a standard of care in other countries (Smaill, 2007).

Of the urine specimens from pregnant women suspected to have UTI, about one-fifth was positive for bacterial cultures with significant bacteriuria. This is similar to the findings reported previously in Ethiopia (Assefa et al., 2008) but slightly higher than that reported by Olsen et al. (2000) and Masinde et al. (2009) in Arusha and Mwanza, Tanzania. Treatment of both SB and ASB in pregnancy can prevent occurrence of adverse outcomes for mother such as septicemia, pyelonephritis, hypertension, pre-eclampsia and premature labour as well as adverse outcome for the unborn child (Raz, 2003).

In this study, Gram negative bacteria were more prevalent (61.9%) than Gram positive bacteria (38.1%). Similar findings have been reported in rural Tanzania (Blomberg et al., 2005) as well as elsewhere (Gebre-Selassie, 1998; Delzell & Lefevre, 2000; Nicolle, 2001; Schnarr & Smaill, 2008), E. coli was major pathogen isolated from the urine cultures and accounted for one-third of the positive cultures with significant bacteriuria. E. coli is considered uropathogenic due to a number of virulence factors specific for colonisation and invasion of the urinary epithelium, such as the P-fimbria and S-fimbria adhesions (Sheffield & Cunningham, 2005). Klebsiella species and coagulase negative Staphylococcus were the second and third most common bacteria isolated.

Although the spectrum of agents causing UTI in pregnant women is relatively constant, their antibiotic susceptibility patterns are different in different geographical locations. The current study showed high level of resistance to first line antimicrobial drugs such as cotrimoxazole. These findings concur with findings from previous studies (Gupta et al., 2001; Arredondo-García et al. 2007; Arredondo-García & Amábilé-Cuevas, 2008).
2008). Most antimicrobials used to treat UTI including cotrimoxazole can achieve high urinary concentrations. This could imply that urinary isolates which were moderately resistant could be effectively treated with normal doses of antibiotics. However, some previous studies have reported that clinical cure rates may be lower among outpatient women with uncomplicated UTI treated with cotrimoxazole when the infecting pathogen is resistant to cotrimoxazole (Masterton & Bochsler, 1995; Talan et al., 2000, Raz et al., 2002). Low level of resistance was found against nitrofurantoin, and this might be due to its narrow range of clinical indications, which results in less usage. Considering the in vitro activity of nitrofurantoin and availability as an oral drug, nitrofurantoin should be considered as one of the drugs of choice against UTI in pregnant women where multi-resistant uropathogens are prevalent. However, the low resistance patterns of nitrofurantoin together with high resistance to cotrimoxazole observed in the current study call for the need to strengthen surveillance to identify changes in sensitivity pattern among urinary tract isolates.

The current study showed high level of cefotaxime resistance. Cefotaxime is second line antimicrobial agent in the third generation of cephalosporins. A possible explanation for the resistance found might be the presence of Extended Spectrum Beta-Lactamase (ESBL) in these strains. Noteworthy to mention that cefotaxime alone may not be enough as a screening drug for ESBL. However, this is an important finding, since it is likely to indicate that UTI in pregnant women at MNH may be present. Similar strains have been reported previously in septicemia patients from the same hospital (Blomberg et al., 2005). It is therefore important that routine screening of ESBL in urinary isolates at MNH is carried out to prevent widespread of resistant isolates. The emergence of ESBLs in community-acquired infections has been reported recently (Arredondo-García & Amábile-Cuevas, 2008). Resistance to ciprofloxacin although relatively low, could have been contributed by an increase in fluoroquinolone prescribing practices for uncomplicated UTI in recent years (Nicolle et al, 2006).

This study is limited by the fact it included highly selected pregnant women with symptoms of UTI only and the duration of the study was only 3-year period. Further studies are recommended to include both symptomatic and asymptomatic pregnant women with UTI over longer duration to establish a trend in antibiotic resistance pattern.

In conclusion, *E. coli* was found to be the common cause of UTI among the pregnant women. Low to moderately high level of resistance against first line drugs and high level of resistance against third generation cephalosporin was observed. It is recommended to monitor the levels of resistance for nitrofurantoin, fluoroquinolone and cefotaxime and screen for Extended Spectrum Beta Lactamase production among cefotaxime resistant *E. coli* and *Klebsiella*.

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